

# Harnessing Amygdalin In Integrative Medicine: Novel Insights For Endocrine Disorders

Vishal Kajla<sup>1</sup>, Bipin Singh<sup>2\*</sup>, Muskaan<sup>3</sup>, Subham Kumar Dogra<sup>4</sup>, Pratibha Sharma<sup>5</sup>, Vishali<sup>6</sup>, Mohd. Sayam<sup>7</sup>, Ajay Bilandi<sup>8</sup>

**ABSTRACT** Amygdalin, often known as vitamin B17, is a cyanogenic glycoside that is mostly present in almond, cherry, and apricot seeds. The FDA has limited its usage in the United States because of its metabolism into hydrogen cyanide, which offers serious health hazards, even though it has historically been used for its alleged anticancer properties. Amygdalin and its derivatives, such as prunasin and mandelonitrile, may have pharmacological qualities, such as antiinflammatory and antioxidant benefits, notwithstanding these reservations. The complex pathophysiology of Polycystic Ovary Syndrome (PCOS), a common endocrine disorder characterized by hyperandrogenism and prolonged anovulation, includes inflammation as a major component in addition to hormonal and metabolic dysfunctions. Common in individuals, insulin resistance is linked to elevated levels of inflammatory markers such TNF- $\alpha$ , CRP, and IL-6. The therapy of PCOS may benefit from the use of interleukin-10 (IL-10), an antiinflammatory cytokine. Due to insulin resistance and other metabolic abnormalities linked to the disorder, PCOS and type 2 diabetes are significantly correlated. This mechanism involves the MAPK/ERK1/2 signaling pathway in addition to the crucial function of GLUT transporters in glucose metabolism. The possibility of cyanide poisoning raises concerns about amygdalin's safety, even though it may have therapeutic advantages for oxidative stress and reproductive health. To investigate its potential advantages while maintaining patient safety, more study is required.

**KEYWORDS:** Amygdalin, Laetrile, Vitamin B17, Cyanide, Anticancer properties, Polycystic Ovary Syndrome (PCOS), Insulin resistance, Chronic inflammation, Type 2 diabetes, Antioxidant properties

## INTRODUCTION

### Amygdalin sources (Vitamin B17):

Over time, proponents of natural medicine have come to recognize amygdalin, commonly known as "laetrile" and "vitamin B17," and research has suggested that it may have anticancer properties [1]. In the past thirty years, this vitamin has generated the greatest controversy. This cyanogenic plant glucoside, which is a member of the Rosaceae family, is discovered in the pits of many fruits and raw nuts, including almonds, cherries, peaches, plums, and apricot stones. Additionally, plants including sorghum, clover, and lima beans contain it [2]. The words laevorotatory and mandelonitrile are the sources of the word laetrile [1,3]. Mandelonitrile refers to a substance's chemical identity, whereas laevorotary defines stereochemistry. The chemical formula for amygdalin is  $C_{20}H_{10}NO_{11}$ , and its estimated molecular weight is 457.42 Dalton. In the past, it was used to cure leprosy, leukoderma, asthma, and bronchitis [1]. Apricot kernels contain a chemical called amygdalin, which emits cyanide. Despite its

dubious effectiveness and FDA warnings about cyanogenic dangers and a lack of clinical proof, amygdalin has been used globally, notably in the treatment of cancer [4]. During the 1920s, Dr. Ernst T. Krebs, Sr., a German scientist, conducted research on apricot kernel extract that contains amygdalin. This compound releases cyanide, which is harmful to humans because of its impact on intestinal bacteria. In 1952, his son, Krebs Jr., created 'laetrile', a substance structurally different from amygdalin. He later renamed it 'Vitamin B-17' in 1970[5]. Early studies focused on the potential anticancer effects of amygdalin and laetrile by examining the involvement of  $\beta$ -glucosidase in cancer cell activity [6]. According to Krebs Jr.'s theory, amygdalin releases more HCN when  $\beta$ -glucosidase levels in cancer cells are higher. Rhodanese, which detoxifies HCN and is present in both cancer and healthy cells, makes amygdalin's selective toxicity against cancer cells more difficult to achieve [7]. The theory put forth by Drs. Krebs and Krebs Jr., according to which glucosidase activity exclusively affects cancer cells, has also been proven to be incorrect because this enzyme has been detected in physiologically normal tissues, albeit in smaller quantities [8,9]. When the vitamin B-17 was given a new name in 1970, it attempted to get around the law prohibiting the use of pharmaceuticals. The usage of vitamins, particularly vitamin B-17, was prohibited [10]. In 1977, 23 states

<sup>1,3</sup>Assistant Professor, College of Pharmacy, RIMT University, Mandi Gobindgarh, Punjab, India-147301, Email: Vishalkajla52@gmail.com

<sup>2,7</sup>College of Pharmacy, RIMT University, Mandi Gobindgarh, Punjab, India-147301

<sup>4</sup>Assistant Professor, School of Pharmaceutical Sciences, RIMT University, Mandi Gobindgarh, Punjab, India-147301

<sup>5</sup>Assistant Professor, JCDM College, Sirsa, Haryana, India-125056

<sup>6</sup>Chandigarh College of Pharmacy, Landran, Mohali, Punjab, India-140307

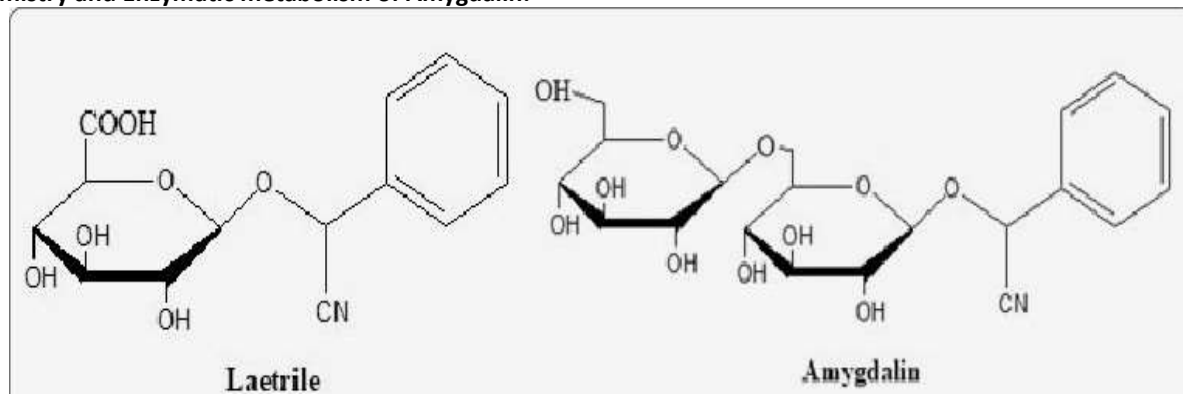
<sup>8</sup>Principal, Bhai Gurdas College of Pharmacy, Sangrur, Punjab

\*Corresponding Authors: Bipin Singh

College of Pharmacy, RIMT University, Mandi Gobindgarh, Punjab, India-147301

were prohibited from receiving laetrile and amygdalin from the U.S. food and drug agency (FDA) [11].

#### Chemistry and Enzymatic metabolism of Amygdalin:



**Figure: 1:** - Biochemical Structural Illustration Coated with laetrile and amygdalin [12].

During the stage of fruit enlargement, amygdalin content typically increases and either stays constant or slightly declines as the fruit ripens. The endocarp of a peach seed has a higher concentration of amygdalin than the mesocarp. The cyanogenic amygdalin diglucoside concentration in almond kernels determines their bitterness [13]. The first step in the production of amygdalin is the conversion of L-phenylalanine to mandelonitrile, which is facilitated by cytochrome P450 and CYP71AN24. The enzyme UDP-glucosyltransferase transforms mandelonitrile into prunasin. Amygdalin is produced via the catalytic conversion of prunasin by the glucosyltransferase [14]. When tissue is macerated or digested, plants that contain cyanogenic glycosides (CGs) release hydrogen cyanide (HCN) through  $\beta$ -glycosidases and hydroxynitrile lyases, which may be inactivated by heat. In plants that lack  $\beta$ -glycosidases, several animals and humans depend on gut endosymbiotes for CG hydrolysis [15]. The bacterial flora in the intestine that can manufacture  $\beta$ -glycosidase in the brush border of the small intestine is likely responsible for the decisive synthesis of HCN in humans [16–18].

Amygdalin, the primary cyanogenic glycoside found in apricot kernels, 59 milligrams of hydrogen cyanide (HCN), which exists in its dissociated form as cyanide,

are released when one gram of amygdalin is broken down. The fact that cyanide is extremely hazardous to people must be noted [19]. When orally consumed, amygdalin undergoes degradation by digestive enzymes during the salivary and gastrointestinal phases, resulting in the formation of prunasin as the main metabolite.  $\beta$ -glucosidase breaks down prunasin further to produce mandelonitrile. Amygdalase and prunase are examples of glucosidase enzymes that generate hydroxy mandelonitrile (149 Da) through hydroxylation across the small intestine wall. It eventually breaks down to produce hydrogen cyanide and benzaldehyde [20–22]. Despite not being hazardous in and of itself, amygdalin is broken down by certain enzymes to form the toxic chemical HCN [23]. According to recent studies, HCN is released in healthy cells, which could be harmful to the body [24]. The harmful effects of cyanide compounds are a result of the breakdown of amygdalin [25,26]. By attaching itself to a ferric ion in the mitochondrial cytochrome oxidase a3, cyanide prevents oxygen from being reduced to water, which inhibits cellular respiration [27]. The primary causes of cyanide poisoning are cellular hypoxia and disruption of aerobic cell metabolism, which lead to cardiovascular and central nervous system dysfunctions [28].

### Pharmacological Activities of Amygdalin's derivatives

**Table: 1** Amygdalin's derivatives with their pharmacological action

Derivative	Pharmacological Action	Reference(s)
<b>Derivative</b>	<b>Pharmacological Action</b>	
<b>Prunasin</b>	Anti-inflammatory, Antioxidant	[29]
<b>Mandelonitrile</b>	Anticancer, Antioxidant	[30]
<b>Amygdalin Hydrolase</b>	Enzyme, enhances amygdalin metabolism	[31]
<b>N-Methyl-βphenylethylamine</b>	Antidepressant, Neuroprotective	[32]
<b>Phenylacetoneitrile</b>	Anticancer, Anti-inflammatory	[33]
<b>Mandelic Acid</b>	Antibacterial, Antifungal	[34]
<b>L-Mandelonitrile</b>	Anticancer, Antioxidant	[35]
<b>Hydrocyanic Acid</b>	Cytotoxic, used in controlled doses for specific treatments	[29]
<b>Benzaldehyde</b>	Anticancer, Antifungal	[34]
<b>Amygdalin-7-O-glucoside</b>	Antioxidant and anti-inflammatory effects	[36]
<b>Amygdalin-2-O-acetate</b>	Cardioprotective effects	[37]
<b>Amygdalin-3-O-glucoside</b>	Anti-diabetic effects	[38]

#### Biosynthesis of amygdalin:

A CYP79 enzyme hydroxylates the amino acid phenylalanine in amygdalin to phenyl acetaldoxime, which a CYP71 enzyme then further hydroxylates to mandelonitrile. The subsequent attachment of one glucose molecule to the α-hydroxyl group of mandelonitrile is catalyzed by a uridine diphosphate glucose-glucosyl transferase (UGT), producing prunasin (d-(-)-mandelonitrile-β-d-glucoside, CAS number 99-18-

3, 295.3 g mol<sup>-1</sup>). When another glucose molecule is added to the 6'-hydroxyl group, it transforms into the di-glucoside gentiobiose, which is then converted to amygdalin (Figure 2). Plants produce amygdalin using the same basic process as CNGs, which involves cytochrome P450 (CYP) enzymes hydroxylating an amino acid to an oxime and then α-hydroxy nitrile, then glycosylating the latter [12].

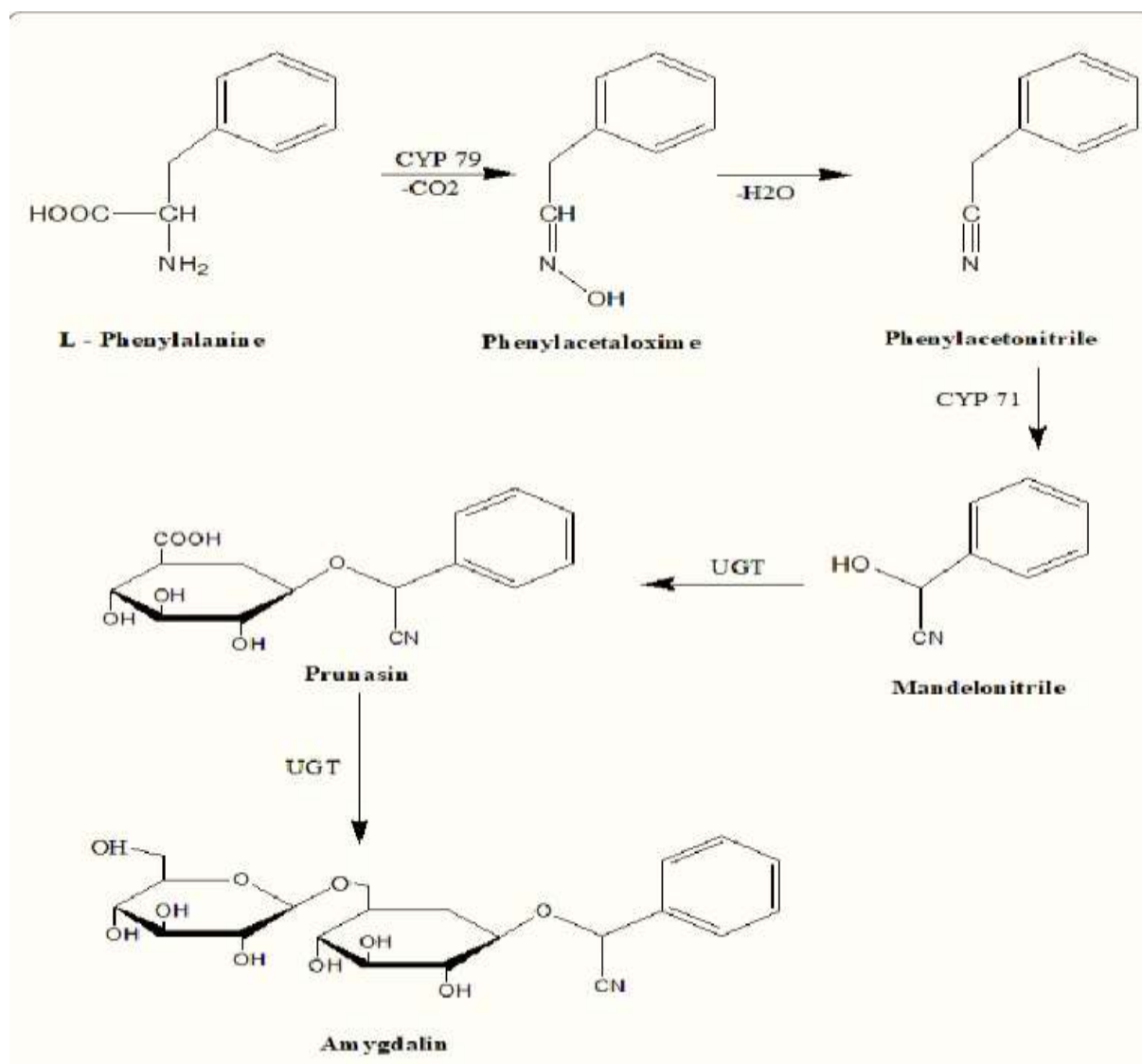


Figure 2: - Amygdalin the process of biosynthesis is shown. [12].

#### Pharmacological Activities of drug in PCOS:

The condition known as The hallmarks of polycystic ovarian syndrome (PCOS) are hyperandrogenism and prolonged anovulation brought on by abnormalities in cellular regulation. Since the initial report by Stein and Leventhal, PCOS has been a subject of controversy in the field of gynecological endocrinology. Our knowledge of the endocrine processes underlying this syndrome's clinical symptoms is currently lacking, disjointed, and frequently unclear [39,40]. PCOS has been observed to run in families, with an autosomal dominant pattern of inheritance, suggesting an oligogenic basis. Several key genes have been implicated in the development of PCOS, including CYP11a, which affects androgen production, and insulin VNTR alleles, which influence hyperinsulinemia and associated risks such as menstrual irregularities and type 2 diabetes mellitus [41]. The pathophysiology of PCOS involves several hypotheses. The LH hypothesis suggests that there is exaggerated LH pulsatility, the insulin hypothesis proposes that insulin resistance leads

to hyperinsulinemia, the ovarian hypothesis suggests excessive ovarian androgen production, and a fourth theory suggests a block in ovarian-level FSH activity despite a normal follicle response to FSH stimulation [42]. The widespread endocrine condition PCOS is not well understood pathophysiologically. It is thought to be the outcome of a convoluted "vicious cycle" with several beginning events that cause ovarian excess androgen production as well as anovulation. A main neuroendocrine malfunction resulting in an increased LH pulse frequency and amplitude; Hyperinsulinemia and resistance to insulin are caused by a specific abnormality in the action and secretion of insulin. An increase in adrenal production of androgenic due to a shift in cortisol metabolism; an increase in ovarian androgen production due to a failure in androgen synthesis [43].

**Anti-Müllerian Hormone (AMH):** Research on AMH and PCOS has been conducted. Granulosa cells in minor antral and preantral follicles produce AMH. AMH levels may be higher in PCOS individuals, particularly in those

who are obese, according to research. In addition to luteinizing hormone (LH) and antral follicle count, AMH can operate as a separate risk factor for the diagnosis of PCOS [44].

**Glucose and Lipid Metabolism:** In PCOS populations, glucose and lipid metabolism have a major impact on hormonal and associated parameters. Particularly in obese people, dysregulation of these metabolic pathways can make PCOS symptoms worse [44].

**The Impact of GnRH in PCOS:** An increasing amount of evidence suggests that increased

GnRH pulse frequency and intensity might promote the synthesis of LH rather than FSH, which would raise the LH/FSH ratio in PCOS-afflicted women [45]. The following data supports the idea that elevated LH levels play a critical role in the development of metabolic and reproductive problems. The first is that LH causes ovarian theca cells to produce more androgen, which results in hyperandrogenism and a stoppage of follicle growth [46]. Second, a higher frequency of LH pulses prevents the generation of FSH and estrogen, which delays ovulation and follicle development. Thirdly, the development of polycystic ovaries in PCOS patients is facilitated by LH, which increases ovarian IGF-1 release, which can improve LH binding and androgen synthesis in theca cells [47].

The etiology of PCOS, a common gynaecological endocrine condition in women of reproductive age, is still unclear. Its development is thought to be significantly influenced by inflammation, though. IL-6, TNF- $\alpha$ , IL-17, CRP, NLR, and PLR are among the inflammatory markers that have been studied in connection with PCOS. TNF- $\alpha$  is an inflammatory cytokine, while IL-6 is a protein of the immune system linked to inflammation. IL-17 has been linked to PCOS, and CRP is a marker produced in response to inflammation. NLR indicates cardiovascular health and metabolic syndrome, while PLR has recently been associated with PCOS. It is noteworthy that the ratio of CRP to albumin can function as an accurate diagnostic for PCOS. Additionally, mean platelet volume (MPV) and PLR are also relevant indicators in this context [48-50].

#### **Inflammatory markers related to PCOS:**

##### **Interleukins and their subtypes:**

##### **IL-6 (Interleukin-6):**

Adipocytes from the body's fat reserves produce a multitude of cytokines, Interleukin 6, also known as IL-6, is among them [51]. PCOS is associated with higher IL-6 levels and are regulated by NF- $\kappa$ B. It is also associated with various diseases such as rheumatoid arthritis, cardiovascular conditions, asthma, and colon cancer, highlighting its dual pro- and antiinflammatory roles [52]. In its normal function, IL-6 supports epithelial renewal and immune function. However, in PCOS, insulin resistance may lead to increased IL-6 levels, indicating potential alterations in immunity and an

increased risk of cardiovascular issues in young women [53]. Nevertheless, persistent inflammation caused by disorders like PCOS can pose significant dangers.

##### **IL-8 (Interleukin-8):**

IL-8, a cytokine that promotes inflammation, functions specifically as an activator and chemoattractant for neutrophils [54]. The role of IL-8 in regulating the ovary shows promise. It plays a part in the maturation of oocytes, ovulation, and follicular development [55]. According to earlier studies, the ovary's granulosa cells, stromal cells, and theca cells all contain IL-8 mRNA [56,57]. Goto et al. [58] have demonstrated that IL-8, as a signaling molecule, is involved in follicular development through vascularization. Research has documented the existence of IL-18 within the follicular fluid of patients undergoing in vitro fertilization (IVF) and females with regular menstrual cycles [59]. Ascites, blood, tumour tissue, and fluids from ovarian cysts in individuals with ovarian cancer have all been found to have elevated IL-8 levels [60,61]. Proliferation, adhesion, invasion, and angiogenesis are all improved by increased IL-8 expression [62]. Ovarian endometrium has increased IL-8 expression [63]. Serum IL-8 levels were linked to neo angiogenesis, metastasis, and melanoma, according to Nicolae et al. [64]. According to recent research, PCOS is associated with elevated IL-8 levels [65]. IL-8 was shown to be high in PCOS women in a clinical trial conducted by Ali et al. [66,67], but it reduced after receiving pioglitazone and metformin.

##### **IL-10 (Interleukin-10):**

The cytokine interleukin-10 (IL-10), which suppresses the immune system and reduces inflammation, is essential to our body's defensive systems [68]. It suppresses the activity of TH1 cells and was first discovered to be a component of TH2 cells [69]. It is thought to support the maintenance of pregnancy by reducing TH1 cell activity, which promotes progesterone production and corpus luteum maturation [70]. A low concentration of IL-10 has been associated with metabolic syndrome and obesity [71]. PCOS patients have shown a reduction in plasma IL-10 levels [72]. According to Sylus et al. [73], clomiphene citrate up-regulates IL10, causing PCOS women to ovulate and become pregnant at higher rates. Predicting the relationship between IL-10 and PCOS is difficult because the majority of research on the subject has been population-based [72,74] and has a limited sample size. The role of IL-10 in the pathophysiology of PCOS requires more investigation.

##### **IL-17 (Interleukin-17):**

Recent research has found a connection between PCOS and IL-17, IL-17a, and IL-1Ra [75–77]. Inflammatory and autoimmune disorders are primarily affected by IL-17a [76]. Higher levels of IL-1Ra in PCOS affect glucose metabolism and lessen insulin resistance. Increases in

inflammatory cytokines (IL-17a, IL-1a, IL-1b, IL-2, and IL-8) can impair ovarian activity, while high levels of AMH are associated with heightened inflammation and metabolic and reproductive dysfunctions [77].

#### **TNF- $\alpha$ (Tumor Necrosis Factor-alpha):**

Adipose tissue secretes TNF- $\alpha$ , IL-6, and adiponectin, with their levels being altered in cases of obesity. Changes in adipokine profiles can lead to a slightly proinflammatory condition that impacts both adipose tissue and other specific tissues [78,79]. TNF- $\alpha$  acts as a proinflammatory cytokine. By preventing the activity of tyrosine kinase of the receptor for insulin in fat and muscle cells, it contributes to the development of systemic resistance to insulin (IR), which is linked to obesity. On the other hand, adiponectin functions as an insulin sensitizer and possesses anti-inflammatory properties [80-82]. The pathophysiology of PCOS is associated with TNF- $\alpha$  due to its role in increasing insulin resistance (IR), causing hyperandrogenism (HA), and influencing follicular formation. It is thought that increased TNF- $\alpha$  expression in muscle tissue and Adipose tissue contributes to the development of IR in people by decreasing the activity of the tyrosine kinase of the insulin receptor [83,84]. Furthermore, TNF- $\alpha$  has been implicated in chronic inflammatory disorders such as ulcerative colitis, rheumatoid arthritis, and Crohn's disease [85-87]. Research has indicated that adiponectin expression may be reduced by TNF- $\alpha$  and IL-6. However, it is worth noting that TNF- $\alpha$  may also have the ability to increase adiponectin expression. In animal models, NF- $\kappa$ B, which is a pathway effector of TNF- $\alpha$ , has been linked to insulin resistance and metabolic disturbances associated with obesity [81]. In the case of PCOS, even in the absence of obesity, there is evidence of a proinflammatory environment that could be influenced by hyperandrogenism. This environment may contribute to the increased production of TNF- $\alpha$  by macrophages [88]. Females with PCOS have a greater amount of CRP (C-reactive protein) than people of the same age and BMI, which may indicate that PCOS has an underlying inflammatory mechanism [89,90]. The secretion of free fatty acid into the circulation by visceral adipocytes in non-obese women with PCOS has been found to assist modestly enhance inflammation in these individuals [81].

#### **CRP (C-reactive protein):**

The underlying cause of long-term cardiovascular hazards in PCOS may be persistent lowgrade inflammation. Obesity and insulin resistance contribute to dysfunction of endothelial cells, stiffness of the arteries, early ventricular alterations, and the onset of atherosclerosis, which may be brought on by oxidative stress, dyslipoproteinemia, hypertension, and disturbed homeostasis [91-93]. An raised white blood cell count, inflammatory cytokines including interleukin-6 and

interleukin-18, and high levels of C-reactive protein (CRP) all point to a persistent state of low-grade inflammation that is characteristic of PCOS [93,94]. Research has indicated that C-reactive protein (CRP) is a very accurate predictor of cardiovascular morbidity and a dependable measure of inflammation [95,90,93], Specifically regarding lipid profile parameters [94]. Kalyan and colleagues discovered that the CRP/albumin ratio is a more accurate marker of inflammation in PCOS compared to free androgens or insulin resistance, demonstrating increased specificity regardless of BMI [96]. In PCOS, inflammation seems to play a significant role regardless of BMI, affected by CRP gene variations, which could be essential in confirming CRP as a reliable predictor of cardiovascular disease [97]. Theoretically, CRP might identify PCOS individuals who are more likely to develop heart disease and type II diabetes [93]. Just two published research articles have examined CRP level in PCOS patients and shown that they were more likely than controls to have increased CRP levels. These studies' authors came to the conclusion that women with PCOS had noticeably increased CRP levels [90,98]. They looked analysed levels of CRP in a larger cohort of PCOS patients who were matched with controls based on BMI in order to either support or refute this link.

#### **NLR (Neutrophil-to-lymphocyte ratio):**

NLR, a marker of inflammation, was utilized in PCOS [99-101]. In PCOS, NLR levels showed an increase even with comparable CRP values [100,101]. Inflammatory haematological indicators such as leucocytes, NLR, and PLR have been connected to the prediction of death from cancer, cardiovascular disorders, and cerebrovascular accidents [102-107]. NLR serves as an indicator of systemic inflammation [105].

#### **PLR (Platelet-to-lymphocyte ratio):**

PLR serves as a biological marker indicating thrombosis-inflammation balance. It correlates with increased megakaryocytic proliferation, thrombocytosis, and high platelet counts, alongside low lymphocyte counts, highlighting inflammation and clotting risks [106]. Activation of platelets and function are correlated with platelet size. Prior research has shown that MPV is a risk factor for cardiovascular illnesses and is a significant predictor of platelet activation [107]. The use of NLR, PLR, and MPV (mean platelet volume) as indicators of chronic inflammation is growing in the literature [105-107]. NLR [99-101] and MPV in PCOS [108-111] have been reported. PLR in PCOS hasn't been reported before, though. As far as we are aware, this article is the first to use NLR, PLR, and MPV in combination for PCOS. Furthermore, no research has looked at the relationship between inflammatory markers and PCOS IVF outcomes.

#### **MAPK/ERK1/2:**



The EMT process is regulated by the MAPK pathway, which is made up of many kinases like as p38, ERK1/2, and JNK [112]. The levels of JNK and p-JNK proteins, as well as the pJNK:JNK ratio, did not significantly differ across the patient groups. According to Western blot analysis, patients with PCOSEH and nPCOSEH showed higher p38 MAPK (MAPK) levels yet did not have p-p38 MAPK levels or p-p38:p38 ratios. Additionally, p-ERK1/2 and the p-ERK1/2:ERK1/2 ratio were greater in PCOS patients than in nPCOS patients, whereas p-ERK1/2 protein expression was also higher in nPCOSEH patients [113]. It has been demonstrated that IRS activation is suppressed by MAPK pathway activation, which encourages the formation of IR [114]. In the case of PCOS, MAPK signaling is a crucial signal transduction route connected to androgen production and IR [115]. Furthermore, decreased GLUT4 expression brought on by MAPK activation may hinder glucose transport [116]. Additionally, there is proof that PI3K/AKT signaling can affect the activation of the MAPK pathway [117]. We

measured MAPK-related protein expression to determine if berberine's effects on IR were connected to MAPK pathway activation. In PCOS model mice, we found a substantial berberine-mediated inhibition of p38, ERK, and JNK. This implies that berberine may modulate the MAPK signaling pathway to attenuate IR in part. In conclusion, our study offers new proof that berberine therapy can reduce IR in a rat model system, hence mitigating the pathophysiology of PCOS through a mechanism most likely associated with GLUT4 overexpression. The molecular processes that underlie this increase are probably connected to the inhibition of MAPK signaling and the stimulation of PI3K/AKT signaling by berberine. Thus, our findings imply that berberine could have therapeutic uses. Crosstalk between MAPK signaling and PI3K-AKT is amply supported by prior research [118]. Our findings support the presence of this type of crosstalk and imply that it might be a major mediator of IR linked to PCOS.

#### GLUT And their Types:

**Table: 2** GLUT Pathway and their role in PCOD and Diabetes:

GLUT TYPE	PATHWAY/ FUNCTION	ROLE IN PCOD	ROLE IN DIABETES	IN REFERENCES
GLUT1	Facilitates glucose uptake in most tissues	Elevated expression in tissues; linked to insulin resistance	Impaired insulin expression contributes to glucose uptake issues	[119]
GLUT2	High capacity glucose transporter in liver, pancreas, kidneys	Implicated in glucose sensing and insulin abnormalities	Dysfunction leads to hyperglycemia and reduced insulin secretion	[120]
GLUT3	Major transporter in neurons, placenta	May affect metabolic pathways influencing ovulation	Essential for glucose uptake in brain; dysfunction can impact cognition and metabolism	[121]
GLUT4	Insulin regulated glucose transporter in muscle and adipose tissue	Insulin resistance common in PCOD may lead to reduced GLUT4 activity	Deficiency linked to insulin resistance and Type 2 Diabetes	[122]
GLUT5	Fructose transporter	Potential link to metabolic syndrome in PCOD	May influence energy metabolism in diabetes	[123]

**JAK-STAT and JKT:****Table: 3** JAK-STAT and JKT (Janus Kinase Tyrosine) signaling Pathway and their role in PCOD and Diabetes:

PATHWAY	ROLE IN PCOS	ROLE IN DIABETES	REFERENCES
JAK-STAT	Inflammation, insulin resistance	Insulin signaling, beta cell function	[124]
JKT	Insulin resistance, metabolic dysfunction	Glucose metabolism, insulin sensitivity	[125]

**NOS And Their Types:****Table: 4** The roles of endothelial nitric oxide synthase (eNOS) and inducible nitric oxide synthase (iNOS) in PCOS and diabetes.

TYPE	ROLE IN PCOS	ROLE IN DIABETES	REFERENCES
iNOS	Inflammation, insulin resistance	Insulin resistance, impaired insulin signaling	[126]
eNOS	Hormonal regulation, endothelial function	Vascular homeostasis, atheroprotection	[127]

**TLRs Receptors and their roles in PCOD and Diabetes:**

The innate immune system depends heavily on a family of proteins called toll-like receptors (TLRs). Immune responses are triggered by their recognition of pathogen-associated molecular patterns (PAMPs) [128].

**Table: 5** TLRs Receptors and their types, their roles in PCOD and Diabetes:

TLR TYPE	ROLE IN PCOS	ROLE IN DIABETES	REFERENCES
TLR2	Inflammation, insulin resistance	Insulin resistance, vascular complications	[129]
TLR4	Inflammation, hormonal imbalance	Insulin resistance, atherosclerosis	[129]
TLR9	Inflammation, metabolic dysfunction	Insulin resistance, diabetic neuropathy	[130]

**PCOS And Diabetes:**

**PCOS** is a hormonal disorder marked by elevated levels of androgen hormones, which contribute to the development of male characteristics. Typical indications encompass the growth of facial hair, irregularity in menstrual cycles, acne, the presence of ovarian cysts, and difficulties with fertility [131].

**Diabetes and Insulin resistance and their relation:**

In women with polycystic ovarian syndrome (PCOS), the link between insulin resistance and hyperandrogenism is well-established [132]. Nevertheless, previous findings [133] indicating an insulin-antagonistic impact of androgens have been overshadowed by more recent research showing that antiandrogen therapy with flutamide [134] or GnRH agonists [135,136] does not modify insulin resistance in PCOS. Conflicting outcomes have been documented in non-PCOS females, with

certain studies [132,137-140] proposing a link between testosterone and insulin resistance, while others [141,142] show no association. Despite this, androgens can impact body composition, which is linked to insulin sensitivity. Therefore, it is plausible that testosterone may indirectly affect insulin sensitivity through its influence on body composition. We present the findings of hormonal, metabolic, and body composition investigations conducted before, as well as 1 month and 9 months after, the removal of a Leydig cell tumor in a postmenopausal woman [143]. The correlation between PCOS and diabetes is significant, although PCOS itself does not directly cause diabetes. Resistance to insulin, a major contributing factor to the development of diabetes type 2, affects both illnesses [144]. PCOS is also linked to resistance to insulin, a condition in which the body fails to use insulin, the hormone that controls blood sugar levels, efficiently.



This illness makes type 2 diabetes more likely to occur [145]. A majority of individuals with PCOS experience some level of insulin resistance, although the exact percentage is unknown. Estimates suggest that approximately 65% to 70% of people with PCOS are insulin-resistant [146]. Insulin resistance and hyperinsulinemia are prevalent in 70% to 80% of individuals with PCOS who have a BMI over 30. However, even those with a lower BMI can be affected. The risk of insulin resistance is more strongly correlated with abdominal fat, indicated by a hip-to-waist ratio of 0.85 or higher, rather than BMI alone [147]. While PCOS may not directly cause diabetes, a study conducted in 2021 discovered that individuals with PCOS have a higher incidence of diabetes. However, this association may be attributed to shared risk factors rather than a direct cause and effect relationship [148]. Whether or not PCOS causes diabetes, it is important to remember that people with PCOS are more likely to get type 2 diabetes. A 2017 study found that body fat levels are linked to this increased risk of diabetes type 2, and that slim PCOS patients are not at a greater risk of developing type 2 diabetes [149]. An autoimmune response that stops insulin synthesis is known as type 1 diabetes [150]. Individuals with type 1 diabetes are at an increased risk of developing PCOS [151]. A comprehensive analysis and meta-analyses from 2016 recommend that individuals with ovaries and type 1 diabetes undergo screening for PCOS due to the heightened risk of developing PCOS and its associated traits [152].

#### **Amygdalin and its Antioxidant effect:**

Lipid production, protein folding, and calcium ion storage are only a few of the cellular functions in which the endoplasmic reticulum (ER) is essential [153]. Numerous illnesses, including overweight and obesity, atherosclerosis, diabetes type 2, hepatic cirrhosis, and renal damage, are strongly associated with ER stress [154]. Studies have demonstrated that disruptions in the ER homeostasis pathway lead to a decrease in very low-density lipoprotein levels, a rise in lipoprotein B100 breakdown and changes to transcription factors linked to lipids [155]. Transcriptional factors and enzymes involved in lipogenesis are overexpressed when ER stress is induced [156]. This can result in inflammation and apoptosis in the hepatic tissue [157]. Recently, The interaction of two cellular stressors, oxidative stress and ER stress, has drawn interest in the study of the pathophysiology of several illnesses [158,159]. Oxidative stress occurs when the body produces too many reactive oxygen species (ROS), which are products of oxygen molecules [160]. ROS can be generated by environmental factors or mitochondrial dysfunction

[161]. Oxidative stress causes apoptotic cascades, releases inflammatory chemicals, and contributes to the development of a number of illnesses, including diabetes, non-alcoholic fatty liver disease, and kidney damage. According to a number of studies, the production of ROS within the ER is facilitated by unfolded proteins and abnormalities in the glutathione (GSH)/GSSH ratio during endoplasmic reticulum (ER) stress [162,163]. According to a research by Kim et al. [164], mice's livers showed reduced GSH levels and enhanced lipid peroxidation as a result of tunicamycin (TM)-induced ER stress. The overexpression of ER stress indicators, on the other hand, is linked to the enhancement of oxidative stress.

The ER stress cascade is triggered by oxidative stress, which also increases the production of C/EBP homologous protein (CHOP), activating transcription factor 4 (ATF4), and activating transcription factor 6 (ATF6) [165,166]. Ali et al [167] demonstrated that supplementation with GSH can effectively alleviate ER stress chaperones. In a separate investigation, selenium, known for its antioxidant properties, was found to downregulate the gene expressions of GRP78, ATF6, and ATF4. The levels of glutathione peroxidase (GSH-Px), catalase (CAT), and superoxide dismutase (SOD) were also shown to increase [168]. The cyanide-containing chemical amygdalin is often found in the seeds of apricots, peaches, bitter almonds, and other rosaceous plants. Because of its ability to block angiogenesis, cure conditions including emphysema, bronchitis, asthma, renal fibrosis, diabetes, and pain, amygdalin has been used historically [169-173].

Moreover, a number of investigations have demonstrated amygdalin's anti-inflammatory and anticancer properties in a range of cell lines and tissues [174–176]. During the first stage of this study, it was shown that amygdalin decreased the lipid profile, ALT, and AST levels in mice's livers during TM-induced ER stress [177]. Amygdalin decreased myeloperoxidase (MPO) and malondialdehyde (MDA) levels after D-galactosamine liver damage, according to a recent research by Tang et al. [178]. The purpose of this study was to assess the effects of amygdalin on oxidant and antioxidant markers in TM-induced ER stress and its relationship to ER stress and oxidative stress, since previous research has not examined the antioxidant qualities of amygdalin in the liver caused by ER stress and its association with oxidative stress. ER stress is linked to various conditions, such as diabetes. The antioxidant effects of amygdalin could potentially alleviate damage associated with ER stress [179].

**Table: 6** The application, model, dosage, key findings in tabular form:

APPLICATION	MODEL/SUBJECT	DOSAGE/Form	KEY FINDINGS	REFERENCE
<b>ANTI-INFLAMMATORY</b>	Various cell lines	Various	reduces pro-inflammatory markers and cytokines to demonstrate antiinflammatory actions.	[180].
<b>ANTICANCER</b>	Cancer cell lines	Various	Induces apoptosis and inhibits proliferation in various cancer cell lines.	[183].
<b>ANTIOXIDANT</b>	Mice Liver	Pre-treatment, Moderate	Reduces MDA, increases SOD, CAT, GSH levels, and reduces oxidative stress.	[180].
<b>DIABETES MANAGEMENT</b>	Animal models	Various	Improves glucose tolerance and reduces blood glucose levels.	[182].
<b>PAIN RELIEF</b>	Various models	Oral, Intramuscular	Provides pain relief by modulating the central and peripheral nervous system.	[181].
<b>ASTHMA AND BRONCHITIS TREATMENT</b>	Animal models	Oral	Reduces inflammation and improves respiratory function in models of asthma and bronchitis.	[183].
<b>HEPATIC PROTECTION</b>	Mice Liver	Pre-treatment, Moderate	Reduces liver damage markers (ALT, AST), improves antioxidant status, and reduces lipid peroxidation.	[180].
<b>RENAL FIBROSIS</b>	Animal models	Various	Alleviates renal fibrosis by modulating oxidative stress and inflammation.	[182].
<b>FEMALE REPRODUCTIVE HEALTH</b>	Animal models	Various	Modulates oxidative balance in reproductive tissues, influences steroidogenesis, and supports reproductive health.	[181].
<b>CARDIOPROTECTIVE EFFECTS</b>	Animal models	Various	Reduces cardiac oxidative stress and improves cardiac function.	[183].

**Cell lines and Targeting: -Amygdalin and their effects especially in PCOS and Diabetes:** In India, the stem and leaves possess medicinal properties for the treatment of menstrual disorders and dysmenorrhea. The plant component contains a phytoconstituent that binds to estrogen receptors and effectively inhibits cell proliferation [184]. Polycystic ovarian disease, an endocrine disorder that causes menstrual irregularities due to hormonal imbalance affecting LH, FSH, estrogen, and testosterone levels, is significantly influenced by insulin resistance [185]. The plant *Scoparia dulcis*, commonly referred to as sweet broom weed, belongs to the Scrophulariaceae family of plants. It can be found in tropical and subtropical regions of India, Myanmar, America, and the West Indies [184,185]. This plant has been traditionally used for various medical purposes, such as managing diabetes, skin issues, kidney stones, menstrual disorders, anti-sickling, anti-cancer, and many other ailments [186]. *S. dulcis* demonstrates hypoglycaemic properties, improving the body's reaction to insulin and maybe helping to treat insulin resistance-related disorders, such as PCOD and ovarian

tumors [187,188]. The aim of this study is to examine the possible anticancer properties of metformin, with a particular emphasis on its use in ovarian cancer (SKOV3 cell line), as well as the identification of plant-derived compounds for further studies on inhibiting cell proliferation, which could be beneficial for PCOD treatment [189]. The chemical composition aided in the successful extraction and identification of *Scoparia dulcis*, as confirmed through analysis of the ethyl acetate extract. The extract's flavonoids and phenolic components were partly responsible for the suppression of SKOV3 ovarian cancer cells. The structural information on these compounds was acquired using HR-LCMS technique [190].

#### **Safety and Side effects:**

**Cyanide Poisoning:** Amygdalin is converted into cyanide within the body, and excessive consumption of amygdalin can result in cyanide poisoning. The following are symptoms of cyanide toxicity: vomiting, headaches, nausea light-headedness, fever, confusion, decreased blood oxygen levels that cause bluish discolouration of the lips and skin, Coma, death, nerve

damage, liver damage, and extremely low blood pressure [191,192].

**Chronic Consumption:** Neuropathy symptoms, such as blurred vision, hearing loss, unsteadiness, and malfunctioning sensory or motor nerves, can be brought on by a long-term diet high in cyanogenic glycosides (such as amygdalin) [191].

### Conclusion:

The relationship between insulin resistance and inflammation is significant when PCOS, a condition characterized by hormonal and metabolic irregularities, is present. Antiinflammatory cytokines like interleukin-10 (IL-10) show potential in treatment options, however research indicates that PCOS patients often have high levels of inflammatory markers such TNF- $\alpha$ , CRP, and IL-6. These two disorders are correlated, which highlights the metabolic challenges faced by PCOS patients, which is related by processes involving the GLUT transporters and the MAPK/ERK1/2 signaling system. Vitamin B17, another name for amygdalin, is a substance that has both serious health hazards and some therapeutic advantages. It has been connected to anticancer effects and is present in seeds. Its conversion into hydrogen cyanide, however, raises questions about safety. The danger of cyanide poisoning cannot be disregarded, even considering amygdalin's possible anti-inflammatory and antioxidant properties. Therefore, further study is necessary to completely understand the compound's safety and efficacy, even if it may offer some therapeutic advantages in treating oxidative stress and reproductive health concerns. This data bolsters the idea that, in order to protect patient safety, any investigation into the advantages of amygdalin must be conducted under strict scientific examination.

### Future prospective:

The study focuses on optimizing the dosage and safety of amygdalin, investigating its antioxidant, anti-inflammatory, and insulin-sensitizing effects, and designing rigorous clinical trials to evaluate its efficacy in treating PCOS and diabetes. It also explores the potential of combining amygdalin with other therapeutic agents to enhance its efficacy and safety profile. Personalized medicine is also explored, focusing on genetic and metabolic factors in individual responses to amygdalin treatment. Alternative derivatives of amygdalin, such as prunasin and mandelonitrile, are being researched to identify compounds with similar therapeutic benefits but lower toxicity. Public awareness and education are also being promoted, emphasizing the importance of controlled dosages and monitoring. Combining amygdalin with Novel Drug Delivery Systems (NDDS) could offer a new approach to managing Polycystic Ovary Syndrome (PCOS), targeting specific pathways while minimizing toxicity risks. Regulatory approvals are being established to ensure

compliance with safety standards for the safe use of amygdalin and its derivatives.

### REFERENCES:

1. Holzbecher MD, Moss MA, Ellenberger HA. The cyanide content of laetrile preparations, apricot, peach and apple seeds. *Journal of Toxicology: Clinical Toxicology*. 1984 Jan 1;22(4):341-7.
2. Do JS, Hwang JK, Seo HJ, Woo WH, Nam SY. Antiasthmatic activity and selective inhibition of type 2 helper T cell response by aqueous extract of semen armeniacae amarum. *Immunopharmacology and Immunotoxicology*. 2006 Jan 1;28(2):213-25.
3. Hwang HJ, Lee HJ, Kim CJ, Shim IS, Hahm DH. Inhibitory Effect of Amygdalin on Lipopolysaccharide-Inducible TNF- $\alpha$  and IL-1 $\beta$  mRNA Expression and Carrageenan-Induced Rat Arthritis. *Journal of microbiology and biotechnology*. 2008;18(10):1641-7.
4. Horneber M, Ernst E, Milazzo S. Laetrile treatment for cancer. *Cochrane Database Syst. Rev*. 2015;2015:4.
5. Liczbiński P, Bukowska B. Molecular mechanism of amygdalin action in vitro: review of the latest research. *Immunopharmacology and immunotoxicology*. 2018 May 4;40(3):212-8.
6. Rauws AG, Olling M, Timmerman A. The pharmacokinetics of amygdalin. *Archives of toxicology*. 1982 Mar;49:311-9.
7. Greenberg DM. The case against laetrile. The fraudulent cancer remedy. *Cancer*. 1980 Feb 15;45(4):799-807.
8. Arafa HM. Possible contribution of  $\beta$ -glucosidase and caspases in the cytotoxicity of glufosfamide in colon cancer cells. *European journal of pharmacology*. 2009 Aug 15;616(13):58-63.
9. Oliveri V, Viale M, Caron G, Aiello C, Gangemi R, Vecchio G. Glycosylated copper (II) ionophores as prodrugs for  $\beta$ -glucosidase activation in targeted cancer therapy. *Dalton Transactions*. 2013;42(6):2023-34.
10. Krebs ET. nitrilosides (vitamin B-17), their nature, occurrence and metabolic significance antineoplastics vitamin B-17. *Journal of applied nutrition*. 1970.
11. Milazzo S, Ernst E, Lejeune S, Boehm K. Laetrile treatment for cancer. *Cochrane Database of Systematic Reviews*. 2006(2).
12. Hoogenboom LA. Scientific opinion: Acute health risks related to the presence of cyanogenic glycosides in raw apricot kernels and products derived from raw apricot kernels. *EFSA Journal*. 2016;14(4):e04424.
13. Lee SH, Oh A, Shin SH, Kim HN, Kang WW, Chung SK. Amygdalin contents in peaches at different

- fruit development stages. Preventive nutrition and food science. 2017 Sep;22(3):237.
14. Del Cueto J, Ionescu IA, Pičmanová M, Gericke O, Motawia MS, Olsen CE, Campoy JA, Dicenta F, Møller BL, Sánchez-Pérez R. Cyanogenic glucosides and derivatives in almond and sweet cherry flower buds from dormancy to flowering. *Frontiers in Plant Science*. 2017 May 19;8:800.
  15. Siegień I. Cyjanogeneza u roślin i jej efektywność w ochronie roślin przed atakiem roślinożerców i patogenów. *Kosmos*. 2007;56(1-2):155-66.
  16. Shim SM, Kwon H. Metabolites of amygdalin under simulated human digestive fluids. *International Journal of Food Sciences and Nutrition*. 2010 Dec 1;61(8):770-9.
  17. Nowak A, Zielińska A. Aktywność przeciwnowotworowa amigdaliny Anticancer activity of amygdalin. *Postępy Fitoter*. 2016;17:282-92.
  18. Rietjens IM, Martena MJ, Boersma MG, Spiegelberg W, Alink GM. Molecular mechanisms of toxicity of important food-borne phytotoxins. *Molecular nutrition & food research*. 2005 Feb;49(2):131-58.
  19. Alexander J, Barregård L, Bignami M, Ceccatelli S, Cottrill B, Edler L, Grasl-Kraupp B, Hogstrand C, Hoogenboom LR, Nebbia CS, Knutsen HK. Acute health risks related to the presence of cyanogenic glycosides in raw apricot kernels and products derived from raw apricot kernels. *Efsa Journal*. 2016.
  20. Shim SM, Kwon H. Metabolites of amygdalin under simulated human digestive fluids. *International Journal of Food Sciences and Nutrition*. 2010 Dec 1;61(8):770-9.
  21. Do JS, Hwang JK, Seo HJ, Woo WH, Nam SY. Antiasthmatic activity and selective inhibition of type 2 helper T cell response by aqueous extract of semen armeniacae amarum. *Immunopharmacology and Immunotoxicology*. 2006 Jan 1;28(2):213-25.
  22. Chang LW, Zhu HP, Li WB, Liu HC, Zhang QS, Chen HB. Protective effects of amygdalin on hyperoxia-exposed type II alveolar epithelial cells isolated from premature rat lungs in vitro. *Zhonghua er ke za zhi= Chinese Journal of Pediatrics*. 2005 Feb 1;43(2):118-23.
  23. Qadir M, Fatima K. Review on pharmacological activity of amygdalin. *Arch Can Res*. 2017;5(4):160.
  24. Liczbiński P, Bukowska B. Molecular mechanism of amygdalin action in vitro: review of the latest research. *Immunopharmacology and immunotoxicology*. 2018 May 4;40(3):212-8.
  25. Lee HM, Moon A. Amygdalin regulates apoptosis and adhesion in Hs578T triple-negative breast cancer cells. *Biomolecules & therapeutics*. 2016 Jan;24(1):62.
  26. Sauer H, Wollny C, Oster I, Tutdibi E, Gortner L, Gottschling S, Meyer S. Severe cyanide poisoning from an alternative medicine treatment with amygdalin and apricot kernels in a 4-year-old child. *Wiener Medizinische Wochenschrift (1946)*. 2015 Jan 22;165(9-10):185-8.
  27. Hamel J. A review of acute cyanide poisoning with a treatment update. *Critical care nurse*. 2011 Feb 1;31(1):72-82.
  28. Coentrão L, Moura D. Acute cyanide poisoning among jewelry and textile industry workers. *The American journal of emergency medicine*. 2011 Jan 1;29(1):78-81.
  29. Tasić-Kostov M, Arsić I, Pavlović D, Stojanović S, Najman S, Naumović S, Tadić V. Towards a modern approach to traditional use: in vitro and in vivo evaluation of *Alchemilla vulgaris* L. gel wound healing potential. *Journal of ethnopharmacology*. 2019 Jun 28;238:111789.
  30. Ma H, Pan JS, Jin LX, Wu J, Ren YD, Chen P, Xiao C, Han J. MicroRNA-17~ 92 inhibits colorectal cancer progression by targeting angiogenesis. *Cancer Letters*. 2016 Jul 1;376(2):293-302.
  31. Motta EV, Gage A, Smith TE, Blake KJ, Kwong WK, Riddington IM, Moran N. Hostmicrobiome metabolism of a plant toxin in bees. *Elife*. 2022 Dec 6;11:e82595.
  32. Zhou X, Teng T, Zhang Y, Del Giovane C, Furukawa TA, Weisz JR, Li X, Cuijpers P, Coghill D, Xiang Y, Hetrick SE. Comparative efficacy and acceptability of antidepressants, psychotherapies, and their combination for acute treatment of children and adolescents with depressive disorder: a systematic review and network meta-analysis. *The Lancet Psychiatry*. 2020 Jul 1;7(7):581-601.
  33. Fan H, Chen L, Sun H, Wang H, Liu Q, Ren Y, Wei D. Development of nitrilase-mediated process for phenylacetic acid production from phenylacetone nitrile. *Chemical Papers*. 2017 Oct;71:1985-92.
  34. Szatraj K, Szczepankowska AK, Chmielewska-Jeznach M. Lactic acid bacteria— promising vaccine vectors: possibilities, limitations, doubts. *Journal of Applied Microbiology*. 2017 Aug 1;123(2):325-39.
  35. Rashid A, Duan X, Gao F, Yang M, Yen A. Roscovitine enhances all-trans retinoic acid (ATRA)-induced nuclear enrichment of an ensemble of activated signaling molecules and augments ATRA-induced myeloid cell differentiation. *Oncotarget*. 2020 Mar 3;11(12):1017.
  36. Zhong O, Hu J, Wang J, Tan Y, Hu L, Lei X. Antioxidant for treatment of diabetic complications: A meta-analysis and systematic review. *Journal of Biochemical and Molecular Toxicology*. 2022 Jun;36(6):e23038.

37. Yang Z, Liu Y, Li Z, Feng S, Lin S, Ge Z, Fan Y, Wang Y, Wang X, Mao J. Coronary microvascular dysfunction and cardiovascular disease: Pathogenesis, associations and treatment strategies. *Biomedicine & Pharmacotherapy*. 2023 Aug 1;164:115011.
38. Chen J, Yi Q, Wang Y, Wang J, Yu H, Zhang J, Hu M, Xu J, Wu Z, Hou L, Zhang Z. Longterm glycemic variability and risk of adverse health outcomes in patients with diabetes: A systematic review and meta-analysis of cohort studies. *Diabetes Research and Clinical Practice*. 2022 Oct 1;192:110085.
39. Stein IF, Leventhal ML. Amenorrhea associated with bilateral polycystic ovaries. *American journal of obstetrics and gynecology*. 1935 Jan 1;29(2):181-91.
40. Insler V, Lunenfeld B. OPINION: Pathophysiology of polycystic ovarian disease: new insights. *Human reproduction*. 1991 Sep 1;6(8):1025-9.
41. Adams JM, Taylor AE, Crowley Jr WF, Hall JE. Polycystic ovarian morphology with regular ovulatory cycles: insights into the pathophysiology of polycystic ovarian syndrome. *The Journal of Clinical Endocrinology & Metabolism*. 2004 Sep 1;89(9):4343-50.
42. Franks S, Gharani N, Waterworth D, Batty S, White D, Williamson R, McCarthy M. Current developments in the molecular genetics of the polycystic ovary syndrome. *Trends in Endocrinology & Metabolism*. 1998 Feb 1;9(2):51-4.
43. Tsilchorozidou T, Overton C, Conway GS. The pathophysiology of polycystic ovary syndrome. *Clinical endocrinology*. 2004 Jan;60(1):1-7.
44. Wang L, Luo M, Yu X, Li R, Ye F, Xiong D, Gong Y, Zheng M, Liu W, Zeng J. Assessing the clinical diagnostic value of anti-Müllerian hormone in polycystic ovarian syndrome and its correlation with clinical and metabolism indicators. *Journal of Ovarian Research*. 2024 Dec;17(1):1-0.
45. Taylor AE, McCourt B, Martin KA, Anderson EJ, Adams JM, Schoenfeld D, Hall JE. Determinants of abnormal gonadotropin secretion in clinically defined women with polycystic ovary syndrome. *The journal of clinical endocrinology & metabolism*. 1997 Jul 1;82(7):2248-56.
46. Gilling-Smith CA, Willis DS, Beard RW, Franks ST. Hypersecretion of androstenedione by isolated thecal cells from polycystic ovaries. *The Journal of Clinical Endocrinology & Metabolism*. 1994 Oct 1;79(4):1158-65.
47. Cara JF, Fan J, Azzarello J, Rosenfield RL. Insulin-like growth factor-I enhances luteinizing hormone binding to rat ovarian theca-interstitial cells. *The Journal of clinical investigation*. 1990 Aug 1;86(2):560-5.
48. Dey R, Bhattacharya K, Basak AK, Paul N, Bandyopadhyay R, Chaudhuri GR, Purkait MP, Bhattacharjee A, Bose C, Shukla N, Bhaduri R. Inflammatory perspectives of polycystic ovary syndrome: role of specific mediators and markers. *Middle East Fertility Society Journal*. 2023 Dec 12;28(1):33.
49. Abraham Gnanadass S, Divakar Prabhu Y, Valsala Gopalakrishnan A. Association of metabolic and inflammatory markers with polycystic ovarian syndrome (PCOS): an update. *Archives of Gynecology and Obstetrics*. 2021 Mar;303:631-43.
50. Ulug E, Pinar AA. A New Approach to Polycystic Ovary Syndrome and Related Cardiometabolic Risk Factors: Dietary Polyphenols. *Current Nutrition Reports*. 2023 Sep;12(3):508-26.
51. Kistner TM, Pedersen BK, Lieberman DE. Interleukin 6 as an energy allocator in muscle tissue. *Nature metabolism*. 2022 Feb;4(2):170-9.
52. Covarrubias AJ, Horng T. IL-6 strikes a balance in metabolic inflammation. *Cell metabolism*. 2014 Jun 3;19(6):898-9.
53. Fulghesu AM, Sanna F, Uda S, Magnini R, Portoghese E, Batetta B. IL-6 serum levels and production is related to an altered immune response in polycystic ovary syndrome girls with insulin resistance. *Mediators of inflammation*. 2011;2011(1):389317.
54. Xie K. Interleukin-8 and human cancer biology. *Cytokine & growth factor reviews*. 2001 Dec 1;12(4):375-91.
55. Arici A, Oral E, Bukulmez O, Buradagunta S, Engin O, Olive DL. Interleukin-8 expression and modulation in human preovulatory follicles and ovarian cells. *Endocrinology*. 1996 Sep 1;137(9):3762-9.
56. Rizk B, Aboulghar M, Smits J, Ron-El R. The role of vascular endothelial growth factor and interleukins in the pathogenesis of severe ovarian hyperstimulation syndrome. *Human Reproduction Update*. 1997 May 1;3(3):255-66.
57. Chang RJ, Gougeon A, Erickson GF. Evidence for a neutrophil-interleukin-8 system in human folliculogenesis. *American journal of obstetrics and gynecology*. 1998 Apr 1;178(4):650-7.
58. Goto J, Suganuma N, Takata K, Kitamura K, Asahina T, Kobayashi H, Muranaka Y, Furuhashi M, Kanayama N. Morphological analyses of interleukin-8 effects on rat ovarian follicles at ovulation and luteinization in vivo. *Cytokine*. 2002 Nov 1;20(4):168-73.
59. Gazvani MR, Bates M, Vince G, Christmas S, Lewis-Jones DI, Kingsland C. Follicular fluid concentrations of interleukin-12 and interleukin-8 in IVF cycles. *Fertility and sterility*. 2000 Nov 1;74(5):953-8.



60. Darai E, Detchev R, Hugol D, Quang NT. Serum and cyst fluid levels of interleukin (IL)6, IL-8 and tumour necrosis factor-alpha in women with endometriomas and benign and malignant cystic ovarian tumours. *Human Reproduction*. 2003 Aug 1;18(8):1681-5.
61. Edgell T, Martin-Roussety G, Barker G, Autelitano DJ, Allen D, Grant PL, Rice GE. Phase II biomarker trial of a multimarker diagnostic for ovarian cancer. *Journal of cancer research and clinical oncology*. 2010 Jul;136:1079-88.
62. Wang Y, Xu RC, Zhang XL, Niu XL, Qu Y, Li LZ, Meng XY. Interleukin-8 secretion by ovarian cancer cells increases anchorage-independent growth, proliferation, angiogenic potential, adhesion and invasion. *Cytokine*. 2012 Jul 1;59(1):145-55.
63. Fasciani A, D'ambrogio G, Bocci G, Monti M, Genazzani AR, Artini PG. High concentrations of the vascular endothelial growth factor and interleukin-8 in ovarian endometriomata. *Molecular Human Reproduction*. 2000 Jan 1;6(1):50-4.
64. Ene Nicolae CD, Nicolae I. Interleukin 8 serum concentration, but not lactate dehydrogenase activity, positively correlates to CD34 antigen in melanoma tumors. *Journal of Immunoassay and Immunochemistry*. 2016 Sep 2;37(5):463-71.
65. Yoshimoto T, Yoshimoto T, editors. *Cytokine frontiers: Regulation of immune responses in health and disease*. Springer Science & Business Media; 2013 Oct 28.
66. Cordero MD, Alcocer-Gómez E, editors. *Inflammasomes: clinical and therapeutic implications*. Springer International Publishing; 2018 Sep 4.
67. Shah M, Ali A, Malik MO, Rehman F, Badshah H, Ehtesham E, Vitale SG. Treatment with metformin and combination of metformin plus pioglitazone on serum levels of IL-6 and IL-8 in polycystic ovary syndrome: a randomized clinical trial. *Hormone and Metabolic Research*. 2019 Nov;51(11):714-22.
68. Moore KW, de Waal Malefyt R, Coffman RL, O'Garra A. Interleukin-10 and the interleukin-10 receptor. *Annual review of immunology*. 2001 Apr;19(1):683-765.
69. Fiorentino DF, Zlotnik A, Mosmann TR, Howard M, O'Garra A. IL-10 inhibits cytokine production by activated macrophages. *Journal of immunology (Baltimore, Md.: 1950)*. 1991 Dec 1;147(11):3815-22.
70. Hashii K, Fujiwara H, Yoshioka S, Kataoka N, Yamada S, Hirano T, Mori T, Fujii S, Maeda M. Peripheral blood mononuclear cells stimulate progesterone production by luteal cells derived from pregnant and non-pregnant women: possible involvement of interleukin-4 and interleukin-10 in corpus luteum function and differentiation. *Human reproduction (Oxford, England)*. 1998 Oct 1;13(10):2738-44.
71. Scarpelli D, Cardellini M, Andreozzi F, Laratta E, Hribal ML, Marini MA, Tassi V, Lauro R, Perticone F, Sesti G. Variants of the interleukin-10 promoter gene are associated with obesity and insulin resistance but not type 2 diabetes in Caucasian Italian subjects. *Diabetes*. 2006 May 1;55(5):1529-33.
72. Talaat RM, Mohamed YA, Mohamad EH, Elsharkawy M, Guirgis AA. Interleukin 10 (- 1082 G/A) and (- 819 C/T) gene polymorphisms in Egyptian women with polycystic ovary syndrome (PCOS). *Meta gene*. 2016 Sep 1;9:254-8.
73. Sylus AM, Nandeesh H, Sridhar MG, Chitra T, Sreenivasulu K. Clomiphene citrate increases nitric oxide, interleukin-10 and reduces matrix metalloproteinase-9 in women with polycystic ovary syndrome. *European Journal of Obstetrics & Gynecology and Reproductive Biology*. 2018 Sep 1;228:27-31.
74. Karadeniz M, Erdogan M, Zengi A, Tamsel S, Berdeli A, Saygili F, Yilmaz C. Polymorphism of the interleukin-10 gene in polycystic ovary syndrome. *International journal of immunogenetics*. 2008 Apr;35(2):119-23.
75. Foroozanfard F, Soleimani A, Arbab E, Samimi M, Tamadon MR. Relationship between IL-17 serum level and ambulatory blood pressure in women with polycystic ovary syndrome. *Journal of nephropathology*. 2017 Jan;6(1):15.
76. Özçaka Ö, Buduneli N, Ceyhan BO, Akcali A, Hannah V, Nile C, Lappin DF. Is interleukin17 involved in the interaction between polycystic ovary syndrome and gingival inflammation?. *Journal of periodontology*. 2013 Dec;84(12):1827-37.
77. Kuang H, Duan Y, Li D, Xu Y, Ai W, Li W, Wang Y, Liu S, Li M, Liu X, Shao M. The role of serum inflammatory cytokines and berberine in the insulin signaling pathway among women with polycystic ovary syndrome. *PLoS One*. 2020 Aug 12;15(8):e0235404.
78. Mitchell M, Armstrong DT, Robker RL, Norman RJ. Adipokines: implications for female fertility and obesity. *Reproduction*. 2005 Nov 1;130(5):583-97.
79. Cao H. Adipocytokines in obesity and metabolic disease. *Journal of endocrinology*. 2014 Feb 1;220(2):T47-59.
80. Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor- $\alpha$ : direct role in obesity-linked insulin resistance. *Science*. 1993 Jan 1;259(5091):8791.
81. Oróstica L, Astorga I, Plaza-Parrochia F, Vera C, Garcia V, Carvajal R, Gabler F, Romero C, Vega M. Proinflammatory environment and role of TNF- $\alpha$  in endometrial function of obese women having



- polycystic ovarian syndrome. *International journal of obesity*. 2016 Nov;40(11):1715-22.
82. Peraldi P, Hotamisligil GS, Buurman WA, White MF, Spiegelman BM. Tumor necrosis factor (TNF)- $\alpha$  inhibits insulin signaling through stimulation of the p55 TNF receptor and activation of sphingomyelinase. *Journal of Biological Chemistry*. 1996 May 31;271(22):13018-22.
  83. Thathapudi S, Kodati V, Erukkambattu J, Katragadda A, Addepally U, Hasan Q. Tumor necrosis factor-alpha and polycystic ovarian syndrome: a clinical, biochemical, and molecular genetic study. *Genetic testing and molecular biomarkers*. 2014 Sep 1;18(9):6059.
  84. Escobar-Morreale HF, Calvo RM, Sancho J, San Millán JL. TNF- $\alpha$  and hyperandrogenism: a clinical, biochemical, and molecular genetic study. *The Journal of Clinical Endocrinology & Metabolism*. 2001 Aug 1;86(8):3761-7.
  85. Gareb B, Otten AT, Frijlink HW, Dijkstra G, Kosterink JG. local tumor necrosis factor- $\alpha$  inhibition in inflammatory bowel disease. *Pharmaceutics*. 2020 Jun 11;12(6):539.
  86. Jang DI, Lee AH, Shin HY, Song HR, Park JH, Kang TB, Lee SR, Yang SH. The role of tumor necrosis factor alpha (TNF- $\alpha$ ) in autoimmune disease and current TNF- $\alpha$  inhibitors in therapeutics. *International journal of molecular sciences*. 2021 Mar 8;22(5):2719.
  87. Pagnini C, Cominelli F. Tumor necrosis factor's pathway in Crohn's disease: potential for intervention. *International journal of molecular sciences*. 2021 Sep 24;22(19):10273.
  88. Figueroa F, Motta A, Acosta M, Mohamed F, Oliveros L, Forneris M. Role of macrophage secretions on rat polycystic ovary: its effect on apoptosis. *Reproduction*. 2015 Nov 1;150(5):437-48.
  89. Duleba AJ, Dokras A. Is PCOS an inflammatory process?. *Fertility and sterility*. 2012 Jan 1;97(1):7-12.
  90. Kelly CC, Lyall H, Petrie JR, Gould GW, Connell JM, Sattar N. Low grade chronic inflammation in women with polycystic ovarian syndrome. *The Journal of Clinical Endocrinology & Metabolism*. 2001 Jun 1;86(6):2453-5.
  91. Yildiz BO, Bozdogan G, Yapici Z, Esinler I, Yarali H. Prevalence, phenotype and cardiometabolic risk of polycystic ovary syndrome under different diagnostic criteria. *Human reproduction*. 2012 Oct 1;27(10):3067-73.
  92. Bates GW. Polycystic ovary syndrome: a reproductive and metabolic web of risk, comorbidities, and disease. *Fertility and Sterility*. 2019 Mar 1;111(3):471-2.
  93. Blumenfeld Z. The possible practical implication of high CRP levels in PCOS. *Clinical Medicine Insights: Reproductive Health*. 2019 Jul;13:1179558119861936.
  94. Ridker PM, Hennekens CH, Buring JE, Rifai N. C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. *New England journal of medicine*. 2000 Mar 23;342(12):836-43.
  95. Boulman N, Levy Y, Leiba R, Shachar S, Linn R, Zinder O, Blumenfeld ZJ. Increased Creactive protein levels in the polycystic ovary syndrome: a marker of cardiovascular disease. *The Journal of Clinical Endocrinology & Metabolism*. 2004 May 1;89(5):2160-5.
  96. Kalyan S, Goshtesabi A, Sarraz S, Joannou A, Almawi WY. Assessing C reactive protein/albumin ratio as a new biomarker for polycystic ovary syndrome: a case-control study of women from Bahraini medical clinics. *BMJ open*. 2018 Oct 1;8(10):e021860.
  97. Hage FG, Szalai AJ. The role of C-reactive protein polymorphisms in inflammation and cardiovascular risk. *Current atherosclerosis reports*. 2009 Mar;11(2):124-30.
  98. Lejman-Larysz K, Pietrzyk D, Ćwiertnia A, Kozłowski M, Kwiatkowski S, Szydłowska I, Nawrocka-Rutkowska J, Brodowski J, Sowińska-Przepiera E, Cymbaluk-Płoska A, Brodowska A. Influence of hsCRP parameter on the occurrence of metabolic syndrome in patients with polycystic ovary syndrome. *Biomedicines*. 2023 Jul 10;11(7):1953.
  99. Keskin Kurt R, Okyay AG, Hakverdi AU, Gungoren A, Dolapcioglu KS, Karateke A, Dogan MO. The effect of obesity on inflammatory markers in patients with PCOS: a BMI-matched case-control study. *Archives of gynecology and obstetrics*. 2014 Aug;290:315-9.
  100. Yilmaz MA, Duran C, Basaran M. The mean platelet volume and neutrophil to lymphocyte ratio in obese and lean patients with polycystic ovary syndrome. *Journal of Endocrinological Investigation*. 2016 Jan;39:45-53.
  101. Agacayak E, Tunc SY, Sak S, Basaranoglu S, Yüksel H, Turgut A, Gul T. Levels of neopterin and other inflammatory markers in obese and non-obese patients with polycystic ovary syndrome. *Medical science monitor: international medical journal of experimental and clinical research*. 2015;21:2446.
  102. Lee CD, Folsom AR, Nieto FJ, Chambless LE, Shahar E, Wolfe DA. White blood cell count and incidence of coronary heart disease and ischemic stroke, and mortality from cardiovascular disease in African-American and white men and women: The atherosclerosis risk in communities study.
  103. Hoffman M, Blum A, Baruch R, Kaplan E, Benjamin M. Leukocytes and coronary heart disease. *Atherosclerosis*. 2004 Jan 1;172(1):1-6.

104. Templeton AJ, Ace O, McNamara MG, Al-Mubarak M, Vera-Badillo FE, Hermanns T, Šeruga B, Ocana A, Tannock IF, Amir E. Prognostic role of platelet to lymphocyte ratio in solid tumors: a systematic review and meta-analysis. *Cancer epidemiology, biomarkers & prevention*. 2014 Jul 1;23(7):1204-12.
105. Balta S, Celik T, Mikhailidis DP, Ozturk C, Demirkol S, Aparci M, Iyisoy A. The relation between atherosclerosis and the neutrophil-lymphocyte ratio. *Clinical and applied thrombosis/hemostasis*. 2016 Jul;22(5):405-11.
106. Balta S, Ozturk C. The platelet-lymphocyte ratio: a simple, inexpensive and rapid prognostic marker for cardiovascular events. *Platelets*. 2015 Oct 3;26(7):680-1.
107. Yuri Gasparyan A, Ayzvazyan L, P Mikhailidis D, D Kitas G. Mean platelet volume: a link between thrombosis and inflammation?. *Current pharmaceutical design*. 2011 Jan 1;17(1):47-58.
108. Dogan BA, Arduc A, Tuna MM, Karakılıç E, Dagdelen I, Tutuncu Y, Berker D, Guler S. Association of mean platelet volume with androgens and insulin resistance in nonobese patients with polycystic ovary syndrome. *International journal of endocrinology and metabolism*. 2014 Oct;12(4).
109. Silfeler DB, Kurt RK, Yengil E, Un B, Arica S, Baloglu A. Evaluation of Mean Platelet Volume values in lean women with polycystic ovary syndrome. *Pakistan Journal of Medical Sciences*. 2014 May;30(3):589.
110. Köşüş N, Köşüş A, Turhan NÖ. Relationship of ovarian volume with mean platelet volume and lipid profile in patients with polycystic ovary syndrome. *Experimental and Therapeutic Medicine*. 2011 Nov 1;2(6):1141-4.
111. Dasanu CA, Clark 3rd BA, Ichim TE, Alexandrescu DT. Polycystic ovary syndrome: focus on platelets and prothrombotic risk. *Southern medical journal*. 2011 Mar 1;104(3):174-8.
112. Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. *Nature reviews Molecular cell biology*. 2014 Mar;15(3):178-96.
113. Hu M, Zhang Y, Li X, Cui P, Li J, Brännström M, Shao LR, Billig H. Alterations of endometrial epithelial-mesenchymal transition and MAPK signaling components in women with PCOS are partially modulated by metformin in vitro. *Molecular Human Reproduction*. 2020 May;26(5):312-26.
114. Hanai Y, Adachi S, Yasuda I, Takai S, Matsushima-Nishiwaki R, Kato H, Enomoto Y, Akamatsu S, Sakakibara S, Ogura S, Iwama T. Collagen-induced p38 MAP kinase activation is a biomarker of platelet hyper-aggregation in patients with diabetes mellitus. *Life sciences*. 2009 Aug 26;85(9-10):386-94.
115. Diamanti-Kandarakis E, Dunaif A. Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. *Endocrine reviews*. 2012 Dec 1;33(6):981-1030.
116. Zhou DN, Li SJ, Ding JL, Yin TL, Yang J, Ye H. MIF may participate in pathogenesis of polycystic ovary syndrome in rats through MAPK signaling pathway. *Current medical science*. 2018 Oct;38(5):853-60.
117. Zhou T, Meng X, Che H, Shen N, Xiao D, Song X, Liang M, Fu X, Ju J, Li Y, Xu C. Regulation of insulin resistance by multiple MiRNAs via targeting the GLUT4 signaling pathway. *Cellular Physiology and Biochemistry*. 2016 May 28;38(5):2063-78.
118. Choi J, Kim KJ, Koh EJ, Lee BY. Gelidium elegans extract ameliorates type 2 diabetes via regulation of MAPK and PI3K/Akt signaling. *Nutrients*. 2018 Jan 6;10(1):51.
119. Lee MH, Yoon JA, Kim HR, Kim YS, Lyu SW, Lee BS, Song H, Choi DH. Hyperandrogenic milieu dysregulates the expression of insulin signaling factors and glucose transporters in the endometrium of patients with polycystic ovary syndrome. *Reproductive Sciences*. 2020 Aug;27:1637-47.
120. Lamy CM, Sanno H, Labouebe G, Picard A, Magnan C, Chatton JY, Thorens B. Hypoglycemia-activated GLUT2 neurons of the nucleus tractus solitarius stimulate vagal activity and glucagon secretion. *Cell metabolism*. 2014 Mar 4;19(3):527-38.
121. Engin AB, Engin ED, Karakus R, Aral A, Gulbahar O, Engin A. N-Methyl-D aspartate receptor-mediated effect on glucose transporter-3 levels of high glucose exposed-SH-SY5Y dopaminergic neurons. *Food and Chemical Toxicology*. 2017 Nov 1;109:465-71.
122. Ezech U, Chen IY, Chen YH, Azziz R. Adipocyte insulin resistance in PCOS: relationship with GLUT-4 expression and whole-body glucose disposal and  $\beta$ -cell function. *The Journal of Clinical Endocrinology & Metabolism*. 2020 Jul;105(7):e2408-20.
123. Hu M, Zhang Y, Guo X, Jia W, Liu G, Zhang J, Li J, Cui P, Sferruzzi-Perri AN, Han Y, Wu X. Hyperandrogenism and insulin resistance induce gravid uterine defects in association with mitochondrial dysfunction and aberrant reactive oxygen species production. *American Journal of Physiology-Endocrinology and Metabolism*. 2019 May 1;316(5):E794-809.
124. Gurzov EN, Stanley WJ, Pappas EG, Thomas HE, Gough DJ. The JAK/STAT pathway in obesity and diabetes. *The FEBS Journal*. 2016 Aug 1;3002-3015. doi: 10.1111/febs.13709
125. Zhao H, Zhang J, Cheng X, Nie X, He B. Insulin resistance in polycystic ovary syndrome across

- various tissues: an updated review of pathogenesis, evaluation, and treatment. *Journal of Ovarian Research*. 2023 Jan 11;16(1):9.
126. Fujimoto M, Shimizu N, Kunii K, Martyn JJ, Ueki K, Kaneki M. A role for iNOS in fasting hyperglycemia and impaired insulin signaling in the liver of obese diabetic mice. *Diabetes*. 2005 May 1;54(5):1340-8.
  127. Sharma A, Sellers S, Stefanovic N, Leung C, Tan SM, Huet O, Granville DJ, Cooper ME, de Haan JB, Bernatchez P. Direct endothelial nitric oxide synthase activation provides atheroprotection in diabetes-accelerated atherosclerosis. *Diabetes*. 2015 Nov 1;64(11):3937-50.
  128. Woolard MD, Kevil CG. Paying the toll for glucose regulation: a central role for TLR3. *Diabetes*. 2015 Oct;64(10):3345.
  129. Jialal I, Kaur H. The role of toll-like receptors in diabetes-induced inflammation: implications for vascular complications. *Current diabetes reports*. 2012 Apr;12:172-9.
  130. Shen J, Dai Z, Li Y, Zhu H, Zhao L. TLR9 regulates NLRP3 inflammasome activation via the NF- $\kappa$ B signaling pathway in diabetic nephropathy. *Diabetology & metabolic syndrome*. 2022 Feb 4;14(1):26.
  131. Zehra B, Khursheed AA. Polycystic ovarian syndrome: symptoms, treatment and diagnosis: a review. *Journal of Pharmacognosy and Phytochemistry*. 2018;7(6):875-80.
  132. BURGHEN GA, GIVENS JR, KITABCHI AE. Correlation of hyperandrogenism with hyperinsulinism in polycystic ovarian disease. *The Journal of Clinical Endocrinology & Metabolism*. 1980 Jan 1;50(1):113-6.
  133. WOODARD TL, BURGHEN GA, KITABCHI AE, WILIMAS JA. Glucose intolerance and insulin resistance in aplastic anemia treated with oxymetholone. *The Journal of Clinical Endocrinology & Metabolism*. 1981 Nov 1;53(5):905-8.
  134. Diamanti-Kandarakis E, Mitrakou A, Hennes MM, Platanissiotis D, Kaklas N, Spina J, Georgiadou E, Hoffmann RG, Kissebah AH, Raptis S. Insulin sensitivity and antiandrogenic therapy in women with polycystic ovary syndrome. *Metabolism*. 1995 Apr 1;44(4):525-31.
  135. Dunaif A, Green G, Futterweit W, Dobrjansky A. Suppression of hyperandrogenism does not improve peripheral or hepatic insulin resistance in the polycystic ovary syndrome. *The Journal of Clinical Endocrinology & Metabolism*. 1990 Mar 1;70(3):699-704.
  136. Lasco A, Cucinotta D, Gigante A, Denuzzo G, Pedulla M, Trifiletti A, Frisina N. No changes of peripheral insulin resistance in polycystic ovary syndrome after long-term reduction of endogenous androgens with leuprolide. *European journal of endocrinology*. 1995 Dec;133(6):718-22.
  137. Kitabchi AE, Imseis RE, Bush AJ, Williams-Cleaves B, Pourmotabbed G. Racial differences in the correlation between gonadal androgens and serum insulin levels. *Diabetes Care*. 1999 Sep 1;22(9):1524-9.
  138. Evans DJ, Hoffmann RG, Kalkhoff RK, Kissebah AH. Relationship of androgenic activity to body fat topography, fat cell morphology, and metabolic aberrations in premenopausal women. *The Journal of Clinical Endocrinology & Metabolism*. 1983 Aug 1;57(2):304-10.
  139. PEIRIS AN, MUELLER RA, STRUVE MF, SMITH GA, KISSEBAH AH. Relationship of androgenic activity to splanchnic insulin metabolism and peripheral glucose utilization in premenopausal women. *The Journal of Clinical Endocrinology & Metabolism*. 1987 Jan 1;64(1):162-9.
  140. Schriock ED, Buffington CK, Hubert GD, Kurtz BR, Kitabchi AE, Buster JE, Givens JR. Divergent correlations of circulating dehydroepiandrosterone sulfate and testosterone with insulin levels and insulin receptor binding. *The Journal of Clinical Endocrinology & Metabolism*. 1988 Jun 1;66(6):1329-31.
  141. Hauner H, Ditschuneit HH, Pal SB, Moncayo R, Pfeiffer EF. Fat distribution, endocrine and metabolic profile in obese women with and without hirsutism. *Metabolism*. 1988 Mar 1;37(3):281-6.
  142. Toscano V, Bianchi P, Balducci R, Guglielmi R, Mangiantini A, Lubrano C, Sclarra F. Lack of linear relationship between hyperinsulinaemia and hyperandrogenism. *Clinical endocrinology*. 1992 Feb;36(2):197-202.
  143. Volpi E, Rasmussen BB, Lieberman SA, Nagamani M, Ferrer DM, Urban RJ, Gilkison CR. The relationships between testosterone, body composition, and insulin resistance: A lesson from a case of extreme hyperandrogenism. *Diabetes care*. 2005 Feb;28(2):429.
  144. Ovalle F, Azziz R. Insulin resistance, polycystic ovary syndrome, and type 2 diabetes mellitus. *Fertility and sterility*. 2002 Jun 1;77(6):1095-105.
  145. Schmidt TH, Khanijow K, Cedars MI, Huddleston H, Pasch L, Wang ET, Lee J, Zane LT, Shinkai K. Cutaneous findings and systemic associations in women with polycystic ovary syndrome. *JAMA dermatology*. 2016 Apr 1;152(4):391-8.
  146. Marshall JC, Dunaif A. Should all women with PCOS be treated for insulin resistance?. *Fertility and sterility*. 2012 Jan 1;97(1):18-22.
  147. Cho WK, Kim H, Lee HY, Han KD, Jeon YJ, Jung IA, Kim SH, Cho KS, Park SH, Jung MH, Suh BK. Insulin resistance of normal weight central obese adolescents in Korea stratified by waist to height

- ratio: results from the Korea National Health and Nutrition Examination Surveys 2008–2010. *International Journal of Endocrinology*. 2015;2015(1):158758.
148. Zhu T, Cui J, Goodarzi MO. Polycystic ovary syndrome and risk of type 2 diabetes, coronary heart disease, and stroke. *Diabetes*. 2021 Feb 1;70(2):627-37.
  149. Rubin KH, Glintborg D, Nybo M, Abrahamsen B, Andersen M. Development and risk factors of type 2 diabetes in a nationwide population of women with polycystic ovary syndrome. *The Journal of Clinical Endocrinology & Metabolism*. 2017 Oct 1;102(10):3848-57.
  150. Centers for Disease Control, Prevention (US), National Center for Chronic Disease Prevention, Health Promotion (US). Division of Diabetes Translation. Take charge of your diabetes. Department of Health and Human Services; 2002.
  151. Yi Y, El Khoudary SR, Buchanich JM, Miller RG, Rubinstein D, Matthews K, Orchard TJ, Costacou T. Women with Type 1 diabetes (T1D) experience a shorter reproductive period compared with nondiabetic women: the Pittsburgh Epidemiology of Diabetes Complications (EDC) study and the Study of Women's Health Across the Nation (SWAN). *Menopause*. 2021 Jun 1;28(6):634-41..
  152. Escobar-Morreale HF, Roldán-Martín MB. Type 1 diabetes and polycystic ovary syndrome: systematic review and meta-analysis. *Diabetes care*. 2016 Apr 1;39(4):639-48.
  153. Muneer A, Khan RM. Endoplasmic reticulum stress: implications for neuropsychiatric disorders. *Chonnam Medical Journal*. 2019 Jan;55(1):8.
  154. Kaneko M, Imaizumi K, Saito A, Kanemoto S, Asada R, Matsuhisa K, Ohtake Y. ER stress and disease: toward prevention and treatment. *Biological and Pharmaceutical Bulletin*. 2017 Sep 1;40(9):1337-43.
  155. Ariyasu D, Yoshida H, Hasegawa Y. Endoplasmic reticulum (ER) stress and endocrine disorders. *International journal of molecular sciences*. 2017 Feb 11;18(2):382.
  156. DeZwaan-McCabe D, Sheldon RD, Gorecki MC, Guo DF, Gansemer ER, Kaufman RJ, Rahmouni K, Gillum MP, Taylor EB, Teesch LM, Rutkowski DT. ER stress inhibits liver fatty acid oxidation while unmitigated stress leads to anorexia-induced lipolysis and both liver and kidney steatosis. *Cell reports*. 2017 May 30;19(9):1794-806.
  157. Zhu Y, Guan Y, Loor JJ, Sha X, Coleman DN, Zhang C, Du X, Shi Z, Li X, Wang Z, Liu G. Fatty acid-induced endoplasmic reticulum stress promoted lipid accumulation in calf hepatocytes, and endoplasmic reticulum stress existed in the liver of severe fatty liver cows. *Journal of dairy science*. 2019 Aug 1;102(8):7359-70.
  158. Dandekar A, Mendez R, Zhang K. Cross talk between ER stress, oxidative stress, and inflammation in health and disease. *Stress responses: methods and protocols*. 2015:205-14.
  159. Ashraf NU, Sheikh TA. Endoplasmic reticulum stress and oxidative stress in the pathogenesis of non-alcoholic fatty liver disease. *Free radical research*. 2015 Dec 2;49(12):1405-18.
  160. Ore A, Akinloye OA. Oxidative stress and antioxidant biomarkers in clinical and experimental models of non-alcoholic fatty liver disease. *Medicina*. 2019 Jan 24;55(2):26.
  161. Elblehi SS, Hafez MH, El-Sayed YS. L- $\alpha$ -Phosphatidylcholine attenuates mercury-induced hepato-renal damage through suppressing oxidative stress and inflammation. *Environmental Science and Pollution Research*. 2019 Mar 1;26(9):9333-42.
  162. Salim S. Oxidative stress and the central nervous system. *Journal of Pharmacology and Experimental Therapeutics*. 2017 Jan 1;360(1):201-5.
  163. Matsunaga D, Sreekumar PG, Ishikawa K, Terasaki H, Barron E, Cohen P, Kannan R, Hinton DR. Humanin protects RPE cells from endoplasmic reticulum stress-induced apoptosis by upregulation of mitochondrial glutathione. *PLoS one*. 2016 Oct 26;11(10):e0165150.
  164. Kim SH, Kwon DY, Kwak JH, Lee S, Lee YH, Yun J, Son TG, Jung YS. Tunicamycin-induced ER stress is accompanied with oxidative stress via abrogation of sulfur amino acids metabolism in the liver. *International journal of molecular sciences*. 2018 Dec 18;19(12):4114.
  165. Lin H, Liu XB, Yu JJ, Hua F, Hu ZW. Antioxidant N-acetylcysteine attenuates hepatocarcinogenesis by inhibiting ROS/ER stress in TLR2 deficient mouse. *PLoS One*. 2013 Oct 2;8(10):e74130.
  166. Mota SI, Costa RO, Ferreira IL, Santana I, Caldeira GL, Padovano C, Fonseca AC, Baldeiras I, Cunha C, Letra L, Oliveira CR. Oxidative stress involving changes in Nrf2 and ER stress in early stages of Alzheimer's disease. *Biochimica et Biophysica Acta (BBA) Molecular Basis of Disease*. 2015 Jul 1;1852(7):1428-41.
  167. Ali I, Liu HX, Zhong-Shu L, Dong-Xue M, Xu L, Shah SZ, Ullah O, Nan-Zhu F. Reduced glutathione alleviates tunicamycin-induced endoplasmic reticulum stress in mouse preimplantation embryos. *Journal of Reproduction and Development*. 2018;64(1):15-24.
  168. Xiao Y, Xu S, Zhao S, Liu K, Lu Z, Hou Z. Protective effects of selenium against zearalenone-induced apoptosis in chicken spleen lymphocyte via an endoplasmic reticulum stress signaling pathway. *Cell Stress and Chaperones*. 2019 Jan 1;24(1):77-89.



169. Guo J, Wu W, Sheng M, Yang S, Tan J. Amygdalin inhibits renal fibrosis in chronic kidney disease. *Molecular Medicine Reports*. 2013 May 1;7(5):1453-7.
170. Hwang HJ, Kim P, Kim CJ, Lee HJ, Shim I, Yin CS, Yang Y, Hahm DH. Antinociceptive effect of amygdalin isolated from *Prunus armeniaca* on formalin-induced pain in rats. *Biological and Pharmaceutical Bulletin*. 2008 Aug 1;31(8):1559-64.
171. Mirmiranpour H, Khaghani S, Zandieh A, Khalilzadeh OO, Gerayesh-Nejad S, Morteza A, Esteghamati A. Amygdalin inhibits angiogenesis in the cultured endothelial cells of diabetic rats. *Indian Journal of Pathology and Microbiology*. 2012 Apr 1;55(2):211-4.
172. Murnane JM, Ritter S. Alloxan-induced glucoprivic feeding deficits are blocked by Dglucose and amygdalin. *Pharmacology Biochemistry and Behavior*. 1985 Mar 1;22(3):40713.
173. Huaping Z, Liwen C, Wenbin LI, Hanchu L. Effect of amygdalin on the proliferation of hyperoxia-exposed type II alveolar epithelial cells isolated from premature rat. *Journal of Huazhong University of Science and Technology [Medical Sciences]*. 2004 Jun;24:223-
174. Chang HK, Shin MS, Yang HY, Lee JW, Kim YS, Lee MH, Kim J, Kim KH, Kim CJ. Amygdalin induces apoptosis through regulation of Bax and Bcl-2 expressions in human DU145 and LNCaP prostate cancer cells. *Biological and Pharmaceutical Bulletin*. 2006;29(8):1597-602.
175. Chen YU, Ma J, Wang F, Hu J, Cui A, Wei C, Yang Q, Li F. Amygdalin induces apoptosis in human cervical cancer cell line HeLa cells. *Immunopharmacology and immunotoxicology*. 2013 Feb 1;35(1):43-51.
176. Makarević J, Rutz J, Juengel E, Kaulfuss S, Reiter M, Tsaur I, Bartsch G, Haferkamp A, Blaheta RA. Amygdalin blocks bladder cancer cell growth in vitro by diminishing cyclin A and cdk2. *PloS one*. 2014 Aug 19;9(8):e105590.
177. Moslehi A, Farahabadi M, Chavoshzadeh SA, Barati A, Ababzadeh S, Mohammadbeigi A. The effect of amygdalin on endoplasmic reticulum (ER) stress induced hepatic steatosis in mice. *The Malaysian journal of medical sciences: MJMS*. 2018 Feb;25(1):16.
178. Tang F, Fan K, Wang K, Bian C. Amygdalin attenuates acute liver injury induced by Dgalactosamine and lipopolysaccharide by regulating the NLRP3, NF- $\kappa$ B and Nrf2/NQO1 signaling pathways. *Biomedicine & Pharmacotherapy*. 2019 Mar 1;111:527-36.
179. Mo FF, Lv BH, An T, Miao JN, Liu JX, Zhang J, Zhang ZY, Ma MH, Yang XY, Zhao DD, Zhang DW. Protective mechanism of punicalagin against endoplasmic reticulum stress in the liver of mice with type 2 diabetes mellitus. *Journal of functional foods*. 2019 May 1;56:57-64.
180. Moslehi A, Komeili-movahed T, Moslehi M. Antioxidant effects of amygdalin on tunicamycin-induced endoplasmic reticulum stress in the mice liver: Cross talk between endoplasmic reticulum stress and oxidative stress. *Journal of Reports in Pharmaceutical Sciences*. 2019 Jul 1;8(2):298-302.
181. Guo X, Qu FX, Zhang JD, Zheng F, Xin Y, Wang R, Li JY, Li HY, Lu CH. Amygdalin and exercise training exert a synergistic effect in improving cardiac performance and ameliorating cardiac inflammation and fibrosis in a rat model of myocardial infarction. *Applied Physiology, Nutrition, and Metabolism*. 2023 Nov 9;49(3):360-74.
182. He XY, Wu LJ, Wang WX, Xie PJ, Chen YH, Wang F. Amygdalin-A pharmacological and toxicological review. *Journal of ethnopharmacology*. 2020 May 23;254:112717.
183. Ali N. Role of vitamin D in preventing of COVID-19 infection, progression and severity. *Journal of infection and public health*. 2020 Oct 1;13(10):1373-80.
184. Oróstica L, García P, Vera C, García V, Romero C, Vega M. Effect of TNF- $\alpha$  on molecules related to the insulin action in endometrial cells exposed to hyperandrogenic and hyperinsulinic conditions characteristics of polycystic ovary syndrome. *Reproductive Sciences*. 2018 Jul;25(7):1000-9.
185. Akre S, Sharma K, Chakole S, Wanjari MB. Recent advances in the management of polycystic ovary syndrome: a review article. *Cureus*. 2022 Aug 4;14(8).
186. Kumar V, Kumar N. Therapeutic Effect of Herbal Medicinal Plants on Polycystic Ovarian Syndrome: A Review. *Asian Journal of Pharmaceutical Research and Development*. 2022;10(6):153-60.
187. Andhalkar S, Chaware V, Redasani V. A review on medicinal plants of natural origin for treatment of polycystic ovarian syndrome (PCOS). *Asian Journal of Pharmaceutical Research and Development*. 2021 Jun 15;9(3):76-81.
188. Paul M, Vasudevan K, Krishnaja KR. *Scoparia dulcis*: A review on its phytochemical and pharmacological profile. *Innoriginal: Int. J. Sci*. 2017 Jul 27;4(4):17-21.
189. Fu YL, Zhang QH, Wang XW, He H. Antidiabetic drug metformin mitigates ovarian cancer SKOV3 cell growth by triggering G2/M cell cycle arrest and inhibition of mTOR/PI3K/Akt signaling pathway. *Eur Rev Med Pharmacol Sci*. 2017 Mar 1;21(5):116975.
190. B. Mymoonbee, M. Sathish, R. Arunkumar, K. Vamsee Krishna, Isolation, Characterization And Ameliorating Effect Of *Scoparia Dulcis* Linn On

Human Ovarian Cancer Cell Line And Protective Effect On PCOD, Int. J. of Pharm. Sci., 2024, Vol 2, Issue 6, 877-892.  
<https://doi.org/10.5281/zenodo.11671532>

191. He XY, Wu LJ, Wang WX, Xie PJ, Chen YH, Wang F. Amygdalin-A pharmacological and toxicological review. Journal of ethnopharmacology. 2020 May 23;254:112717.
192. Salama RH, Ramadan AE, Alsanory TA, Herdan MO, Fathallah OM, Alsanory AA. Experimental and therapeutic trials of amygdalin. Int. J. Biochem. Pharmacol. 2019 Oct 28;1:21-6.